Thermoregulatory and Cardiac Responses of Infant Spontaneously Hypertensive and Wistar-Kyoto Rats to Cold Exposure

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Abstract—Cardiovascular function during cold exposure is dependent on effective thermoregulation. This dependence is particularly apparent in infants. For example, we have previously demonstrated that in infant rats during cold exposure, cardiac rate is directly related to their ability to produce heat endogenously. The primary source of endogenous heat production for infant rats is brown adipose tissue (BAT). Because of the dependence of cardiac rate on effective thermoregulation in the cold and because hypertension in spontaneously hypertensive rats (SHR) is influenced by the preweanling environment, in this study we examined the thermoregulatory and cardiac rate responses of infant SHR and Wistar-Kyoto rats (WKY) to varying levels of cold exposure. In experiment 1, 7- to 8-day-old SHR and WKY were acclimated at a thermoneutral air temperature (35°C) and then exposed to successive decreases in ambient temperature (30.5°C, 26.5°C, 23°C, and 17°C) while thermal and metabolic measures were recorded. Although both strains increased BAT thermogenesis and oxygen consumption in response to cold exposure, SHR cooled more than WKY and exhibited lower levels of oxygen consumption at the lowest air temperatures. Experiment 2 was identical to experiment 1 except that cardiac rate was also measured. Again, SHR exhibited substantial thermoregulatory deficits compared with WKY; in addition, they were less able than WKY to maintain cardiac rate at the 2 lowest air temperatures tested. Finally, in experiment 3, infant SHR exhibited diminished BAT thermogenesis in response to a range of doses of a selective β3-adrenoceptor agonist. We hypothesize that long-term thermoregulatory deficits during the early postnatal period influence cardiovascular function and contribute to the development of hypertension in SHR. (Hypertension. 1999;33:1465-1469.)

Key Words: body temperature regulation ▪ brown fat ▪ nonshivering thermogenesis ▪ heart rate ▪ rats, inbred strains ▪ hypertension, experimental

The degree of high blood pressure expressed in adult spontaneously hypertensive rats (SHR) is influenced by the neonatal environment.1–3 Thus, manipulations limited to the preweanling period, such as cross-fostering,2 pharmacological interventions,1,4 and handling,5,6 result in the lowering of blood pressure in adult SHR. Such findings highlight the importance of epigenetic processes in the development of established hypertension in SHR.

We have recently demonstrated that thermoregulatory mechanisms play an important role in the protection of cardiovascular function during cold exposure in infant rats.7–9 Specifically, heat produced by brown adipose tissue (BAT) during moderate cold exposure helps to warm cardiac muscle and by doing so contributes to the maintenance of cardiac rate. In contrast, pronounced bradycardia is produced by either extreme air temperatures that overwhelm the ability of BAT to deliver warm blood to cardiac muscle or ganglionic blockade that prevents the activation of BAT thermogenesis in response to the cold.

The ability of BAT thermogenesis to protect cardiac rate during moderate cooling and the loss of this function in response to further decreases in ambient temperature have important implications for cardiovascular control in the infant rat. Because cardiac rate serves as the primary mechanism for the control of cardiac output in infant mammals,10 bradycardia in response to cooling would be associated with either a drop in arterial pressure or an increase in vascular resistance to maintain pressure. Recordings of arterial pressure from infant rats during cold exposure suggest that the latter possibility is more likely, because blood pressure is maintained at basal levels during both moderate and extreme ambient temperatures.8 Thus, it seems that nonshivering thermogenesis buffers against bradycardia during moderate cooling, but as cardiac rate decreases during extreme cooling, vascular resistance must be increased to maintain blood pressure.

Despite the accepted influence of the preweanling environment on the development of hypertension in SHR, the
contribution of thermal factors has not been examined. Therefore, the present study was performed to determine whether thermoregulatory and cardiac responses of infant SHR to cold challenge, exhibiting deficiencies in their ability to maintain cardiac rate as air temperature decreases. This deficiency of SHR may be the result of a number of factors, including reduced body size and, as shown here, diminished BAT thermogenesis. These results suggest that infant SHR are more susceptible to episodes of acute hypothermia and bradycardia that may, as a result, contribute to their development of hypertension.

Methods

Experimental Subjects

Infant male WKY and SHR, born to breeding stock received from Taconic Farms (Germantown, NY) and maintained in our laboratory, were used for the present studies. On the day after birth (day 1), litters were culled to 8 pups and litters with fewer than 6 pups were excluded from the study. All pups were 7 to 8 days old on the day of testing. For experiment 1, 8 WKY pups from 8 litters and 8 SHR pups from 8 litters were used. For experiment 2, 5 WKY pups from 5 litters and 5 SHR pups from 5 litters were used. For experiment 3, 40 WKY pups from 12 litters and 40 SHR pups from 12 litters were used, with littersmates always assigned to different experimental groups. All animals were maintained in polypolyene cages (48 x 20 x 26 cm) with wood shavings as bedding material. Food and water were available ad libitum and a 12:12-hour light-dark cycle was maintained with lights on at 6 AM.

Procedures

The procedures used in the present study have been fully described elsewhere and are only briefly presented here. For experiment 1, an individual pup was removed from a litter on the day of testing, weighed, and placed in an incubator maintained at 35°C to 36°C. Two chromel-constantan thermocouples (Omega) were then secured to the skin surface with collodion as an adhesive. One thermocouple was placed above the BAT in the interscapular region (Tis), and the second was placed in the lumbar region (Tback) distant from the BAT. For experiment 2, pups were lightly anesthetized with ether (~1-minute exposure), and then 3 ECG leads were implanted transcutaneously and secured with collodion. Thermocouples were then placed on the pups as in Experiment 1.

For testing, an individual pup was placed inside a double-walled glass metabolic chamber (height = 17.5 cm; ID = 12.5 cm) maintained at 35°C, a thermoneutral air temperature for 1-week-old rats.11 Air temperature in the chamber (Tan) was controlled by circulation of temperature-controlled water from a water bath through the walls of the chamber. Animals were allowed a 45-minute adaptation period before baseline measurements were acquired for 10 minutes. Tis was then successively decreased to 30°C, 27°C, 23°C, and 17°C, with 45 minutes between each decrease.

Data were acquired with a customized data acquisition system. Thermocouple signals underwent cold-junction compensation before acquisition by the computer. Oxygen consumption (Vo2) was determined with a 2-channel electrochemical oxygen analyzer that compared the oxygen concentration of the airstream passing to the chamber with the exhaust airstream passing from the chamber. The percent difference in oxygen concentration between these 2 air streams was computed to 0.001% and entered in the data acquisition system which computed oxygen consumption in mL O2 kg body wt-1 min-1. For both experiments, thermal and oxygen consumption measurements were recorded twice per minute during the test sessions.

For experiment 2, the ECG signal was amplified and filtered before it was digitized by a second data acquisition system. The ECG analog signal was digitized at a rate of 1000 samples per second, and interbeat intervals (IBI) were determined continuously online with a customized software program with a peak threshold detector (this method provides results identical to direct measurement of intervals between successive R waves).9 IBI data were acquired by the computer at a rate of 30 samples per minute.

For experiment 3, thermocouples were attached in the interscapular and lumbar regions, and the pup was placed in a temperature-controlled chamber to acclimate for 45 minutes at 35°C. After this acclimation period, data collection began for a baseline period of 5 minutes, after which the pup was injected with 0, 0.1, 1.0, 10, or 100 mg/kg of the selective β-adrenoceptor agonist (CL-316243-2; donated by Wyeth-Ayerst, St Davids, Pa) dissolved in isotonic saline. Pups were injected subcutaneously with a volume of 1 μL/kg body wt. Data collection continued for 60 minutes with Tan maintained at 35°C.

Data Analysis

Thermal, metabolic, and cardiac data were imported into StatView 4.5 for the Macintosh. For each pup in experiments 1 and 2, a single score for each variable at each air temperature was determined from measurements taken during the final 2 minutes at each temperature. For experiment 2, cardiac rate values in bpm were determined from the interbeat interval for each pup during this same time period. For experiment 3, maximal change from baseline in Tis and V O2 for each pup during the 60-minute test. A 2-factor ANOVA was used to test the main effects of strain and dose on these variables, and post hoc t tests were used to compare successive air temperatures within a strain and unpaired t tests to compare the strains at each air temperature.

For experiment 3, the maximal change from baseline in Tis, and Tan=Tback was determined for each pup during the 60-minute test. A 2-factor ANOVA was used to test the main effects of strain and dose on these variables, and post hoc t tests were used to compare the 2 strains at each dose.

For all tests, the α level was set at 0.05, and the Bonferroni adjustment was used to correct for multiple post hoc comparisons. All data are presented as mean±SEM.

Results

Body weights were significantly different between strains in experiment 1 (WKY: 17.8±1.2 g; SHR: 12.4±0.9 g; t8=3.5, P<0.005), experiment 2 (WKY: 19.8±0.4 g; SHR: 12.1±0.3 g; t8=15.3, P<0.0001), and experiment 3 (WKY: 22.3±0.4 g; SHR: 13.6±0.5 g; t8=14.0, P<0.0001).

Figure 1 presents Tis and V O2 for the 1-week-old WKY and SHR in experiment 1. The Tis responses to cold exposure were significantly different between infant WKY and SHR (F4,52=30.7, P<0.0001). At the thermoneutral Tis of 35°C, Tis did not differ between the strains, but as Tan decreased, Tis decreased further in SHR than in WKY (F4,52=10.2, P<0.0001). Similarly, the Vo2 responses to cold exposure differed between the 2 strains (F4,52=5.7, P<0.05). Although both strains showed progressive increases in Vo2 as Tis decreased, the WKY pups showed a continued increase in oxygen consumption as Tis decreased to 23°C and maintained oxygen consumption at the Tis of 17°C (F4,52=6.4, P<0.0005). These differences in Vo2 reflect differences in heat production between the 2 strains and are consistent with the greater ability of WKY to maintain Tis during cold exposure.

Figure 2 presents Tis and Vo2 for the infant WKY and SHR in experiment 2. Overall, the thermal and metabolic responses of pups in experiment 2 mirrored those found for pups in experiment 1. Specifically, Tis differed significantly between the strains (F4,32=28.1, P<0.001), with greater decreases in SHR at the lower Tis (F4,32=47.6, P<0.0001). For Vo2,
although there was not a significant main effect of strain (F 1,8 = 0.5), there was a significant strain×Ta interaction (F 4,32 = 25.1, P < 0.0001). Although both strains showed significant increases in VO₂ in response to the initial decrease in Ta, there was a more pronounced decrease in VO₂ for SHR at the lowest Ta.

Cardiac rate for the infant WKY and SHR in Experiment 2 are also presented in Figure 2. There was an overall difference in cardiac rate between WKY and SHR (F 1,8 = 15.6, P < 0.005). Cardiac rate of SHR was elevated at the thermoneutral Ta compared with WKY, consistent with previous findings.12 The 2 strains, however, showed markedly different cardiac rate responses to cooling (F 4,32 = 36.3, P < 0.0001): whereas WKY increased or maintained cardiac rate down to a Ta of 23°C, the cardiac rate of SHR declined steadily, such that at the Ta of 23°C, cardiac rate had fallen significantly and was lower than that of WKY.

Because infant rats cannot shiver,13 increases in VO₂ during cold exposure reflect heat production by BAT. In addition to VO₂, increases in the differential between Ta (measured adjacent to the interscapular BAT) and T_back (measured at a site distant from the interscapular BAT) reflect selective heat production by interscapular BAT as well as heat retention within the thoracic cavity. Figure 3 presents this differential, Ta−T_back, plotted against Ta for the infant WKY and SHR in experiments 1 and 2. These plots indicate an increase in values of Ta−T_back with a decrease in Ta, suggestive of an increase in BAT thermogenesis. In addition, the best-fit polynomial regressions, consistent with the data in Figures 1 and 2, indicate higher values of Ta−T_back for WKY than SHR in both experiments. These strain differences should be interpreted with caution, because the larger body size of the WKY may have influenced its magnitude. Regardless, the patterns of the regression lines are generally consistent with the conclusion that, although both WKY and SHR increased BAT thermogenesis in the cold, WKY were better able to maintain this heat production at the lowest Ta's tested here.

The dose-related thermogenic responses of infant SHR and WKY to the selective β₃ adrenoceptor agonist in experiment 3 are presented in Figure 4. The left panel depicts the maximum changes in Ta to the drug during each 60-minute test. A 2-factor ANOVA indicated significant main effects of dose (F 4,70 = 85.8, P < 0.0001) and strain (F 1,70 = 44.8, P < 0.0001), as well as a significant interaction (F 4,70 = 7.6, P < 0.0001). The right panel depicts the maximum changes in Ta−T_back during each 45-minute test. Again, a 2-factor ANOVA indicated significant main effects of dose (F 4,70 = 48.8, P < 0.0001) and strain (F 1,70 = 19.8, P < 0.0001), as well as a significant interaction (F 4,70 = 4.8, P < 0.005).

Discussion

At a thermoneutral air temperature, cardiac rate was elevated in 1-week-old SHR compared with 1-week-old WKY, al-
though thermal and metabolic measures were equivalent between the strains. In contrast, during cold challenge, SHR were deficient in their thermoregulatory capabilities, with the strain differences becoming more pronounced at the lowest air temperatures tested. Thus, SHR exhibited lower values of $T_{is}$ and oxygen consumption at air temperatures of $\approx 23^\circ C$. These decreased values of $T_{is}$ suggest a diminished capability of SHR to deliver warmed blood to cardiac muscle,7,9 thus leading to the pronounced bradycardia seen in experiment 2.

The diminished thermoregulatory capabilities of infant SHR during cold exposure may be due to exaggerated heat loss and/or insufficient heat production. Because of their smaller size, SHR have larger surface-to-volume ratios versus age-matched WKY, which results in increased heat loss.14 Although the smaller body size of SHR places them at a thermoregulatory disadvantage versus WKY, it may also be true that SHR are deficient in their ability to use BAT to produce heat, as has been shown in adult rats after central injections of prostaglandin E$_2$.15 Indeed, the results of experiment 3 provide direct support for this possibility in that selective activation of BAT with a $\beta_3$-adrenoceptor agonist produced diminished thermogenesis in infant SHR versus age-matched WKY. This diminished thermogenesis was evident at moderate doses of the $\beta_3$-agonist. In contrast, equivalent thermogenesis between the 2 strains was found at the highest dose administered. In other words, the dose response curve of the infant SHR was shifted to the right. These results are consistent with a decrease in sensitivity of the $\beta_3$ adrenoceptor population in BAT of infant SHR, although other factors acting between receptor activation and thermogenesis may be involved. Therefore, these results suggest that both the ability to produce and the ability to retain heat in the cold make the isolated infant SHR more susceptible to hypothermia-induced bradycardia than the isolated WKY.

When the dam is out of the nest, infant rats huddle with littermates, a behavior that results in increased heat retention and metabolic savings in the cold.16 Although the benefits of huddling to each individual pup are considerable, huddling does not replace the need for endogenous heat production at typical colony room air temperatures (G.S., M.S.B., M.M. Adams, written observations, 1999). This raises the question of whether the thermoregulatory deficiencies of individual SHR translate into thermoregulatory deficiencies of huddling SHR. Again, research in our laboratory suggests that this is possible: 1-week-old golden hamsters lack the ability to produce heat endogenously and gain little thermoregulatory advantage from huddling (G.S. and M.S.B., unpublished data, 1998). Thus, even huddling SHR are probably experiencing more frequent and more severe thermal challenges than huddling WKY.

The present results do not support an increased level of sympathetic drive that controls BAT thermogenesis during either basal conditions (ie, thermoneutral) or in response to cold challenge. Similarly, Smith17 found no evidence of
increased sympathetic tone of the levator palpatris muscle in developing SHR versus WKY. These results stand in marked contrast to those that concern sympathetic control of cardiovascular function in preweanling SHR. For example, sympathetic drive of the vasculature is elevated in anesthetized infant SHR, and pressor responses to the sympathetic agonist, methoxamine, are exaggerated. Increased sympathetically mediated pressor responses have also been demonstrated in unanesthetized preweanling SHR. The pressor response of SHR pups to suckling is double that of WKY, and the response can be eliminated by ganglionic blockade. In addition, greater sympathetic influences on cardiac regulation are found in infant SHR. Together, these results suggest that there is not a generalized increase in sympathetic drive of peripheral tissues in SHR during the preweanling period but that the level of sympathetic tone is tissue-specific, with a selective increase in the drive of tissues associated with the cardiovascular system.

Long-term exposure of adult rats to an air temperature of 5°C leads to the development of hypertension within 1 to 2 weeks. Given this finding in adults, the effects of long-term cold exposure and other factors on hypertension would be expected to be much greater in infant rats that are engaged in the process of calibrating various physiological systems. For an infant SHR, bouts of acute cold exposure that produce hypothermia and bradycardia would necessitate increases in peripheral resistance to maintain arterial blood pressure. Increased sympathetically mediated pressor responses have also been demonstrated in unanesthetized preweanling SHR. The pressor response of SHR pups to suckling is double that of WKY, and the response can be eliminated by ganglionic blockade. In addition, greater sympathetic influences on cardiac regulation are found in infant SHR. Together, these results suggest that there is not a generalized increase in sympathetic drive of peripheral tissues in SHR during the preweanling period but that the level of sympathetic tone is tissue-specific, with a selective increase in the drive of tissues associated with the cardiovascular system.

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