

The Development of Day-Night Differences in Sleep and Wakefulness in Norway Rats and the Effect of Bilateral Enucleation

Andrew J. Gall, William D. Todd, Baisali Ray, Cassandra M. Coleman, and Mark S. Blumberg¹

*Program in Behavioral and Cognitive Neuroscience, Department of Psychology,
University of Iowa, Iowa City*

Abstract The suprachiasmatic nucleus exhibits circadian rhythmicity in fetal and infant rats, but little is known about the consequences of this rhythmicity for infant behavior. Here, in experiment 1, the authors measured sleep and wakefulness in rats during the day and night in postnatal day (P)2, P8, P15, and P21 subjects. As early as P2, day-night differences in sleep-wake activity were detected. Nocturnal wakefulness began to emerge around P15 and was reliably expressed by P21. The authors hypothesized that the process of photic entrainment over the 1st postnatal week, which depends on the development of connectivity between the retinohypothalamic tract (RHT) and the SCN, influences the later emergence of nocturnal wakefulness. To test this hypothesis, in experiment 2 infant rats were enucleated bilaterally at P3 and P11, that is, before and after photic entrainment. Whereas pups enucleated at P11 and tested at P21 exhibited increased wakefulness at night, identical to sham controls, pups enucleated at P3 and tested at P21 exhibited the opposite pattern of increased wakefulness during the day. Pups tested at P28 and P35 exhibited this same pattern of increased daytime wakefulness. All together, these results suggest that prenatal and postnatal experience modulates the development of species-typical circadian sleep-wake patterns. Moreover, the authors suggest that visual system stimulation, via the RHT's connections with the SCN, exerts an organizational influence on the developing circadian system and, consequently, contributes to the emergence of nocturnality in this species.

Key words circadian rhythm, nocturnal, diurnal, EMG, development, retinohypothalamic tract, suprachiasmatic nucleus

The circadian system in mammals is regulated in part by an internal "clock" located within the suprachiasmatic nucleus of the hypothalamus (Moore, 1983; Rusak and Zucker, 1979). Endogenous SCN rhythms are first detected prenatally in rats (Reppert et al., 1988). During the early postnatal period in rats, physiological (e.g., metabolic rate) and behavioral (e.g.,

suckling) circadian rhythms are present (Ellison et al., 1972; Henning and Gisel, 1980; Kittrell and Satinoff, 1986; Nuesslein-Hildesheim et al., 1995; Spiers, 1988). However, in comparison to the vast literature on the circadian rhythms of sleep and wakefulness in adults (Fuller et al., 2006), very little is known about such rhythms in infants.

1. To whom all correspondence should be addressed: Mark S. Blumberg, Department of Psychology, University of Iowa, Iowa City, IA, 52242; e-mail: mark-blumberg@uiowa.edu.

On the basis of electroencephalographic measures of sleep, it has been reported that nocturnal wakefulness is expressed by postnatal day 20 (P20) in rats (Frank and Heller, 1997). Because EEG activity cannot be used to provide reliable measures of sleep and wakefulness before P10 (Gramsbergen, 1976), we have relied on electromyographic (EMG) measures instead (Karlsson and Blumberg, 2005). Accordingly, here we use nuchal EMG to assess the development of sleep and wakefulness during the day and night at P2, P8, P15, and P21. We find that the nuchal EMG is sufficiently sensitive to detect day-night differences in sleep-wake activity soon after birth, as well as the emergence of nocturnal wakefulness around P15.

In early infancy—before P8 in rats—the dam entrains her pups' circadian rhythms. Thereafter, light becomes the predominant entraining stimulus (Duncan et al., 1986; Ohta et al., 2002; Takahashi and Deguchi, 1983). This transition from maternal to photic entrainment has been attributed to the development of the retinohypothalamic tract (RHT). Even at birth in rats, melanopsin-containing retinal ganglion cells are directly responsive to light and form functional connections with the SCN via the RHT (Hannibal and Fahrenkrug, 2004; Sekaran et al., 2005). The strength of connectivity between the RHT and SCN increases over the 1st postnatal week and becomes adult-like by the end of the 2nd postnatal week (Hannibal and Fahrenkrug, 2004; Sph and Moore, 1993).

Here, in experiment 2, we examined the role of visual system stimulation on the later emergence of nocturnal wakefulness by enucleating infant rats at P3 and P11, that is, before and after photic entrainment, respectively. Pups enucleated at P3 were tested at P8, P15, P21, P28, and P35, and pups enucleated at P11 were tested at P21. Surprisingly, enucleation at P3 resulted in the development of longer wake bout durations during the *day*, a pattern opposite to the pattern expressed by sham controls as well as pups enucleated at P11. Based on these results, we suggest that visual system stimulation via the RHT during a sensitive period exerts an organizational influence on the development of circadian sleep-wake activity.

MATERIALS AND METHODS

All experiments were performed in accordance with National Institutes of Health guidelines for the care of animals in research and were approved by the Institutional Animal Care and Use Committee of the University of Iowa. All efforts were made to minimize the number of animals used.

Subjects

A total of 264 Sprague-Dawley rats from 64 litters were used. Males and females were equally represented in all experiments, and littermates were always assigned to different experimental groups. Litters were culled to 8 pups within 3 d of birth (day of birth = P0). Mothers and their litters were housed and raised in standard laboratory cages (48 cm long × 20 cm wide × 26 cm high) in the animal colony at the University of Iowa. Food and water were available *ad libitum*. All animals were maintained on a 12-h light-dark schedule with lights on at 0700 h.

Surgery

Implantation of Nuchal EMG Electrodes

In experiments 1 and 2, under isoflurane anesthesia, 2 bipolar stainless steel electrodes (50 μ m diameter, California Fine Wire, Grover Beach, CA) were inserted bilaterally into the nuchal muscles and secured with flexible collodion (Seelke et al., 2005). After surgery, pups recovered in a humidified test chamber maintained at thermoneutrality. All pups were tested on the same day that EMG electrodes were implanted. Regardless of whether testing occurred during the day or night, electrode implantation took place at least 1 h before testing. At night, all surgeries were performed under dim red light illumination, with care being taken to ensure that infants were not exposed to white light.

Bilateral Enucleation

In experiment 2, pups were enucleated at 2 ages: P3 ($n = 160$) and P11 ($n = 32$). For each litter, 4 littermates were enucleated and the remaining 4 received sham surgeries. Under isoflurane anesthesia, each eyelid was opened, the eyeball was removed, and the optic nerve and blood vessels were cauterized. The eyelids were then closed and sealed using Vetbond (3M, St. Paul, MN). The sham surgery was identical up to the point of eyelid opening. Pups recovered in a humidified incubator maintained at thermoneutrality (35 °C) for 3 h. After recovery, all 8 littermates were returned to the home cage and dam until testing.

Apparatus

P2-3 (hereafter P2) and P8-9 (hereafter P8) subjects were tested in an electrically shielded double-walled glass chamber sealed with a rubber lid (height, 18 cm; i.d., 9 cm). P15-16 (hereafter P15), P21-22 (hereafter

P21), P28-29 (hereafter P28), and P35-36 (hereafter P35) subjects were tested in a larger chamber (height, 18 cm; i.d., 12 cm). Air temperature inside each chamber was controlled by regulating the temperature of the water that passed through its walls. Air temperature was regulated at thermoneutrality for each age (i.e., P2: 35.5 °C; P8: 35 °C; P15, P21: 32 °C; P28, P35: 29 °C). Compressed, humidified air passed through the chamber at the rate of 300 ml/min. A round platform constructed of polyethylene mesh was fitted inside each chamber so that the pup could move freely on the platform's surface. The mesh allowed for the movement of air from the bottom of the chamber (where it entered) to the top of the chamber (where it was drawn for analysis of its oxygen content).

Oxygen consumption was measured as described elsewhere (Blumberg and Stolba, 1996). Briefly, a 2-channel electrochemical oxygen analyzer (Ametek, Pittsburgh, PA) was used to compare the oxygen concentration of the air passing through the chamber with that of the nonrespired air. The difference in percentage oxygen between the chamber's effluent airstream and the nonrespired airstream reflects oxygen consumption by the pup. Specifically, by knowing the air flow rate (i.e., 300 ml/min) and the pup's body weight, oxygen consumption can be calculated. All oxygen consumption values are presented as ml O₂/100 g/min.

Nuchal electrodes were connected to differential amplifiers (A-M systems, Carlsborg, WA), and their signals were amplified ($\times 10$ k) and filtered (300-5000 Hz). EMG and oxygen consumption data were acquired (1000 samples/s) and stored using a data acquisition system (BioPac Systems, Santa Barbara, CA).

Procedure

Experiment 1: Development of Day-Night Differences in Sleep and Wakefulness

Seventy-two P2, P8, P15, and P21 male and female rats from 24 litters ($n = 6$ at each age $\times 4$ age groups $\times 3$ test times) were used. Three same-sex littermates were tested successively during the day (hereafter day 1), that night, and the following day (hereafter day 2). Three additional same-sex littermates were tested at one of the 3 other ages. Males and females were equally represented in each age group. Littermates were always assigned to different groups, and no more than 6 pups were tested from a given litter.

On day 1, the first of 3 littermates was implanted with nuchal EMG electrodes and acclimated to the test chamber for at least 1 h, at which time collection

of EMG and oxygen consumption data began. Data acquisition continued for 2 h at P2 and P8 from 1200 to 1400 h, for 3 h at P15 from 1130 to 1430 h, and for 6 h at P21 from 1000 to 1600 h. A second littermate was implanted with nuchal EMG electrodes (again, under red light illumination) and tested that evening. Tests occurred at P2 and P8 from 2400 to 0200 h, at P15 from 2330 to 0230 h, and at P21 from 2200 to 0400 h. On day 2, the third littermate was tested identically to the subject on day 1. Upon completion of testing, each pup was killed with an overdose of sodium pentobarbital.

Experiment 2: Effects of Bilateral Enucleation on Day-Night Differences in Sleep and Wakefulness

One hundred ninety-two male and female rats from 40 litters were used. For subjects receiving enucleation or sham surgery at P3, testing occurred at P8 ($n = 32$), P15 ($n = 32$), P21 ($n = 32$), P28 ($n = 32$), or P35 ($n = 32$). For subjects receiving enucleation or sham surgery at P11, testing occurred only at P21 ($n = 32$). At each age, 2 littermates (one enucleate and one sham) were tested during the day and 2 more were tested at night (i.e., $n = 8$ subjects per group). Males and females were equally represented in each age group. For those subjects tested at P28 and P35, enucleated and sham littermates were weaned at P21 and housed with same-sex siblings in groups of 4.

For pups that had surgery on P3, and on the day of testing at P8, P15, P21, P28, and P35, 2 same-sex littermates, one enucleated and one sham, were implanted with nuchal EMG electrodes and acclimated in separate test chambers for at least 1 h, at which time data collection began for both pups. Data acquisition continued for 2 h at P8 from 1200 to 1400 h, and for 6 h at P15, P21, P28, and P35 from 1000 to 1600 h. At night, 2 additional same-sex littermates were implanted with EMG electrodes (again, under red light illumination) and tested identically to those during the day. Nighttime tests at P8 occurred from 2400 to 0200 h and at P15, P21, P28, and P35 from 2200 to 0400 h. The nighttime tests preceded the daytime tests in half of the subjects. The remaining 4 pups in each litter were tested at one of the two other ages. Finally, for sham and enucleated pups whose surgery occurred at P11 and were tested at P21, the test procedure was identical.

Data Analysis

For all subjects, EMG data were analyzed off-line using BioPac Software. The EMG signal was integrated and full-wave rectified, and then dichotomized into

periods of high muscle tone and atonia (or wakefulness and sleep, respectively), as described earlier (Karlsson et al., 2004). Data were scored by an experienced individual blind to experimental condition. In experiment 2, subjects were occasionally excluded from analysis due to poor or incomplete EMG records. In addition, one pup in experiment 2 was excluded from analysis because its mean sleep duration was greater than 2 standard deviations from the group mean.

Sleep and wake bout durations were imported into Statview 5.0 (SAS Institute, Cary, NC) for analysis. Mean bout durations were determined. Percentage time awake was calculated by dividing the mean wake bout duration by the sum of the mean sleep and wake bout durations, and multiplying by 100. Mean number of cycles per hour was calculated by summing the total number of sleep and wake bouts, dividing by 2, and dividing by the length of the test (h). Mean oxygen consumption (ml O₂/100 g/min) for each pup was calculated from 5 random 10-s periods during sleep.

In experiment 1, a 2-factor analysis of variance was used to test for differences across age and time (i.e., day, night). Planned comparisons using paired *t* tests were used to assess day-night differences within ages. In experiment 2, overall ANOVAs were performed to test for differences across age (for pups enucleated at P3), groups (i.e., sham, enucleate), and time (i.e., day, night). Planned comparisons using unpaired *t* tests were used to assess day-night differences within groups.

Survivor distributions of sleep and wakefulness were produced from each pup and also from pooled data at each age, as described elsewhere (Blumberg et al., 2005). Regression analyses were performed to assess the degree of fit (*r*²) to exponential and power-law distributions.

In addition to log-survivor analyses of pooled wake bout durations, differences between groups were tested by determining, for each P21 subject, the median wake bout duration (*T*_{50%}) and the wake bout duration at the 10th percentile (*T*_{10%}). Group differences in the means of these values were tested using the Mann-Whitney *U* test.

For all tests, alpha was set at 0.05. Means are presented with their standard errors.

RESULTS

Experiment 1: Development of Day-Night Differences in Sleep and Wakefulness

Mean sleep bout durations were significantly lower at night than during the day at P2, P8, and P21

(Figure 1A). ANOVA revealed significant main effects of age ($F_{3,60} = 328.8, P < 0.0001$) and time ($F_{2,60} = 18.3, P < 0.0001$), but not a significant age \times time interaction ($F_{6,60} = 2.2$). For mean wake bout durations (Figure 1B), ANOVA revealed a significant main effect of age ($F_{3,60} = 34.0, P < 0.0001$) and an age \times time interaction ($F_{6,60} = 4.0, P < 0.005$), but not a significant effect of time ($F_{2,60} = 3.0$). Only at P21 were mean wake bout durations significantly different at night than during the day.

For mean percentage of time awake (Figure 1C), ANOVA revealed significant main effects of age ($F_{3,60} = 15.1, P < 0.0001$) and time ($F_{2,60} = 7.7, P < 0.005$), and an age \times time interaction ($F_{6,60} = 2.4, P < 0.005$). At P15 and P21, the mean percentage of time awake was significantly greater at night than during the day.

For mean number of cycles per hour (Figure 1D), ANOVA revealed significant main effects of age ($F_{3,60} = 221.7, P < 0.0001$) and time ($F_{2,60} = 20.7, P < 0.0001$), and an age \times time interaction ($F_{6,60} = 8.7, P < 0.0001$). At P2 and P8, the number of cycles per hour was significantly greater at night than during the day.

In general, mean oxygen consumption during sleep was greater at night than during the day at all 4 ages (Figure 1E). ANOVA revealed main effects of age ($F_{3,60} = 20.1, P < 0.0001$) and time ($F_{2,60} = 12.5, P < 0.0001$), but not a significant age \times time interaction ($F_{6,60} = 0.7$).

Survivor distributions for pooled sleep and wake bout durations are presented in Figure 2. These plots can be used to track the statistical distributions of bout durations across development, as shown previously (Blumberg et al., 2005). In Figure 2A, the sleep bout durations at each age, and during the day and night, fall along a straight line, indicative of an exponential distribution. The trend across development of increasing sleep bout durations is represented on these plots as a decrease in slope with increasing age. Finally, consistent with Figure 1A, sleep bout durations were shorter at night than during the day, particularly at P8, but also at P2 and P21.

As shown in Figure 2B, wake bout durations exhibited pronounced developmental changes in day-night differences and statistical distribution. At P2 and P8, wake bout durations fall along a straight line, again indicative of an exponential distribution; in addition, bout durations were shorter at night than during the day, especially at P2. At P15 and P21 during the day and night, wake bout durations no longer fall along a straight line. Finally, consistent with Figures 1B and 1C, at P15 and even more clearly at P21, wake bout durations were longer at night than during the day, as indicated by the increased percentage of bouts exhibiting longer bout durations.

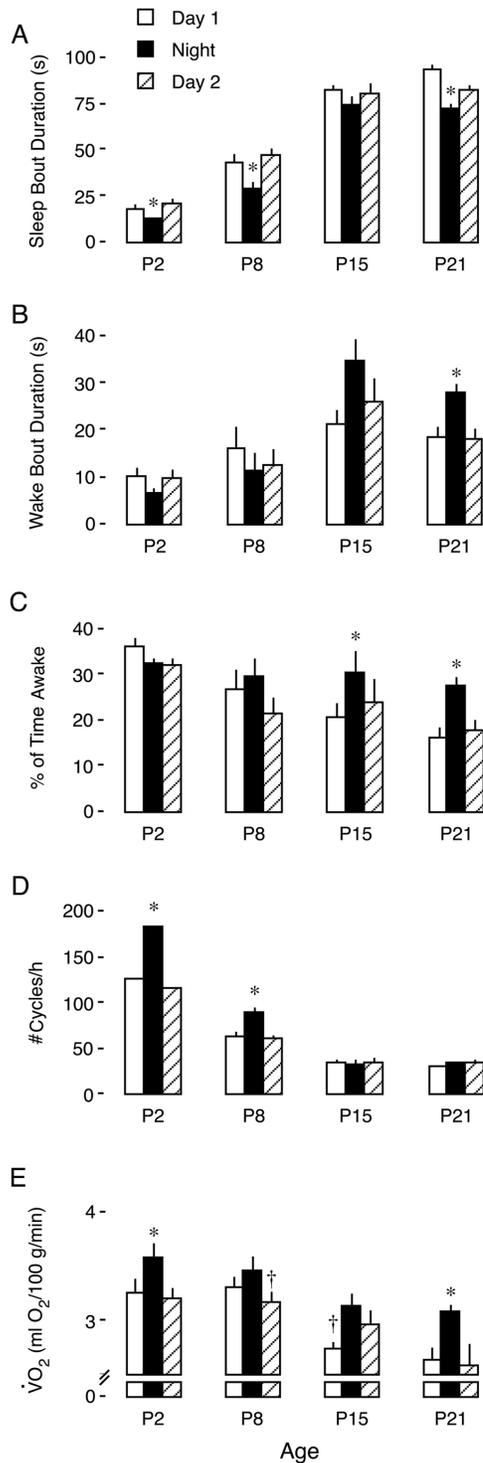


Figure 1. Day-night differences in sleep, wake, and metabolic parameters in infant rats. Mean sleep (A) and wake (B) bout durations during the day and night in P2, P8, P15, and P21 rats. (C) Mean percentage of time spent awake at each age. (D) Mean number of sleep-wake cycles per hour. (E) Mean oxygen consumption determined during periods of sleep. *Night value is significantly different from both day 1 and day 2. †Day 1 or day 2 value is significantly different from night. $n = 6$ subjects per group. Means \pm SE.

We next quantified the fit (r^2) of each subject's sleep and wake bout durations during the day and night to exponential and power-law distributions. For sleep bout durations, ANOVAs performed separately for day 1, night, and day 2 revealed significant main effects of age ($F_{3,40} > 5.5$, $P_s < 0.005$) and distribution ($F_{1,40} > 586.2$, $P_s < 0.0001$), and age \times distribution interactions ($F_{3,40} > 3.9$, $P_s < 0.05$). Paired t tests revealed that sleep bout durations were always better fit by an exponential distribution than by a power-law distribution.

For wake bout durations, ANOVAs performed separately for day 1, night, and day 2 revealed significant main effects of age ($F_{3,40} > 9.8$, $P_s < 0.0001$) and distribution ($F_{1,40} > 30.1$, $P_s < 0.0001$), and age \times distribution interactions ($F_{3,40} > 43.2$, $P_s < 0.0001$). Paired t tests revealed that wake bout durations at P2, regardless of time, were better fit by an exponential distribution than by a power-law distribution, but that this pattern had reversed by P15.

Experiment 2: Effects of Bilateral Enucleation on Day-Night Differences in Sleep and Wakefulness

Of the 96 enucleates, 4 pups died before testing (4.2%); of the 96 shams, 1 pup died before testing (1.0%). In addition, body weights did not differ significantly between enucleates and shams at any age tested. Therefore, we found no evidence that enucleation prevented pups from competing successfully with sham siblings for maternal care or interfered with normal growth.

For mean sleep bout durations and for those subjects that had surgery at P3 (Figure 3; top row), an overall ANOVA revealed a significant age effect ($F_{4,124} = 5.7$, $P < 0.0005$) and significant group \times time ($F_{1,124} = 10.8$, $P < 0.005$) and age \times group \times time ($F_{4,124} = 3.2$, $P < 0.05$) interactions. In contrast, for those subjects that had surgery at P11 and tested at P21, ANOVA did not reveal significant main effects or interactions.

For mean wake bout durations and for those subjects that had surgery at P3 (Figure 3; middle row), an overall ANOVA revealed a significant age effect ($F_{4,124} = 9.3$, $P < 0.0001$) and significant group \times time ($F_{1,124} = 24.9$, $P < 0.0001$) and age \times group \times time ($F_{4,124} = 3.5$, $P < 0.01$) interactions. In contrast, for those subjects that had surgery at P11 and tested at P21, ANOVA revealed only a significant effect of time ($F_{1,24} = 14.4$, $P < 0.001$).

The pattern described above for mean wake bout duration was nearly identical for mean percentage of time awake (Figure 3; bottom row). An overall ANOVA revealed a significant age effect ($F_{4,124} = 4.4$, $P < 0.005$)

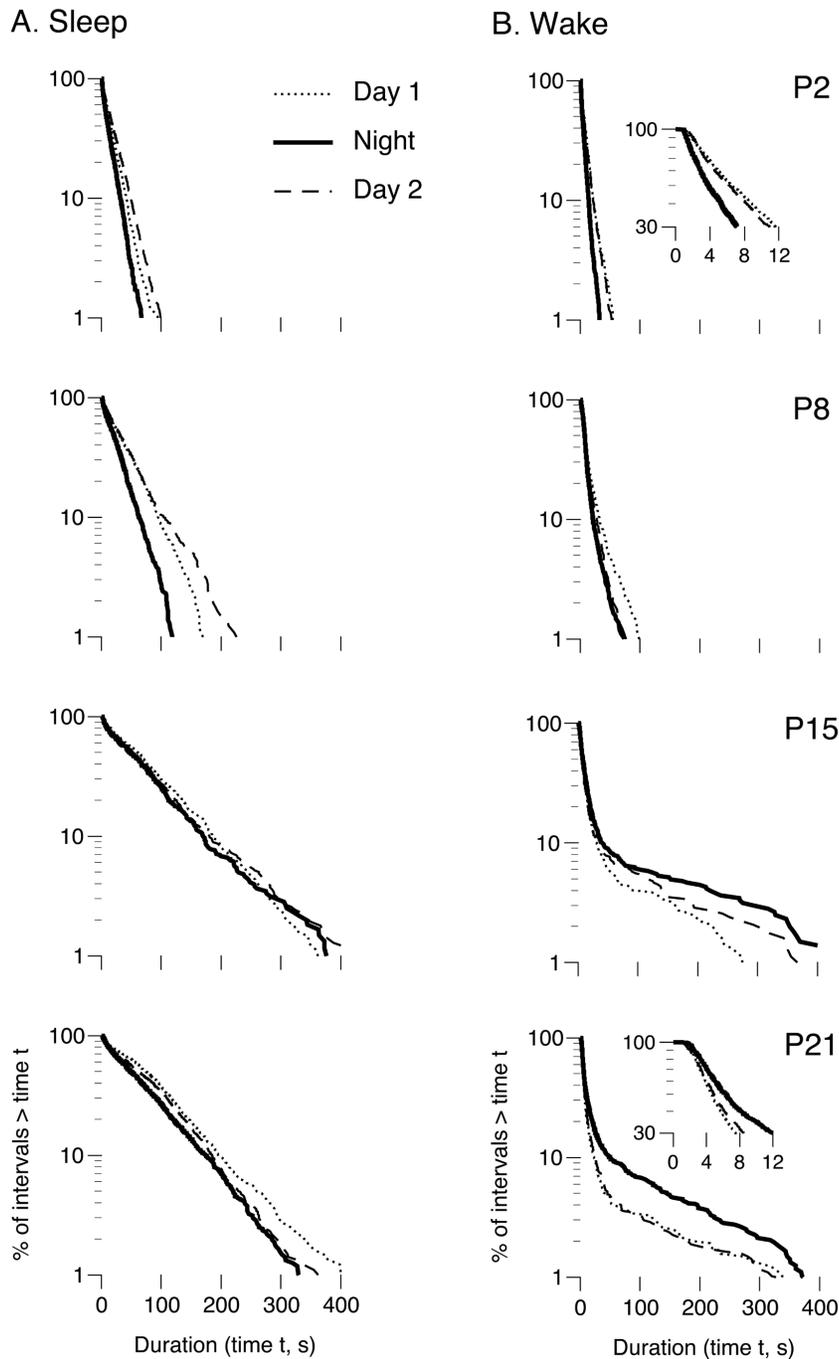


Figure 2. Log-survivor distributions of sleep and wake bout durations. Log-survivor plots of (A) sleep and (B) wake bout durations for rats at P2, P8, P15, and P21 on day 1 (dotted line), at night (solid line), and on day 2 (dashed line). Each plot is constructed from pooled data (620-2213 points per plot). Straight lines on these semi-log plots indicate that the data follow an exponential distribution. Insets provide magnified views at shorter bout durations.

and significant group \times time ($F_{1,124} = 36.6$, $P < 0.0001$) and age \times group \times time ($F_{4,124} = 5.6$, $P < 0.0005$) interactions. Finally, for those subjects that had surgery at P11

and tested at P21, ANOVA again revealed only a significant effect of time ($F_{1,24} = 20.2$, $P = 0.0001$).

Log-survivor plots of pooled P21 wake bout durations for enucleated and sham subjects, presented in Figure 4, reinforce the results described above. For pups that had surgery at P3 (Figure 4A), wake bout durations were longer at night than during the day for the sham pups, whereas the opposite pattern was found for enucleated pups. For pups that had surgery at P11 (Figure 4B), wake bout durations were longer at night than during the day for both the sham and enucleated pups.

Next, for each subject, we determined the wake bout duration at the 10th percentile ($T_{10\%}$). The group means of these values are presented in the right panels of Figure 4. For pups that had surgery at P3 (Figure 4A), sham subjects exhibited longer wake bout durations at night ($z = 2.6$; $P < 0.01$), whereas enucleated subjects exhibited longer wake bout durations during the day ($z = 2.3$; $P < 0.05$). For pups that had surgery at P11 (Figure 4B), both sham ($z = 2.1$, $P < 0.05$) and enucleated ($z = 2.9$, $P < 0.005$) subjects exhibited longer wake bout durations at night. Similar results were obtained for values at $T_{50\%}$ (i.e., median; data not shown).

We found no evidence that enucleation at P3 or P11 differentially affected the statistical distributions of sleep or wake bout durations at any age (data not shown).

DISCUSSION

This study examined the organization of daytime and nighttime sleep-wake activity in rats across early development. Experiment 1 demonstrated that day-night differences in sleep-wake activity can be detected soon

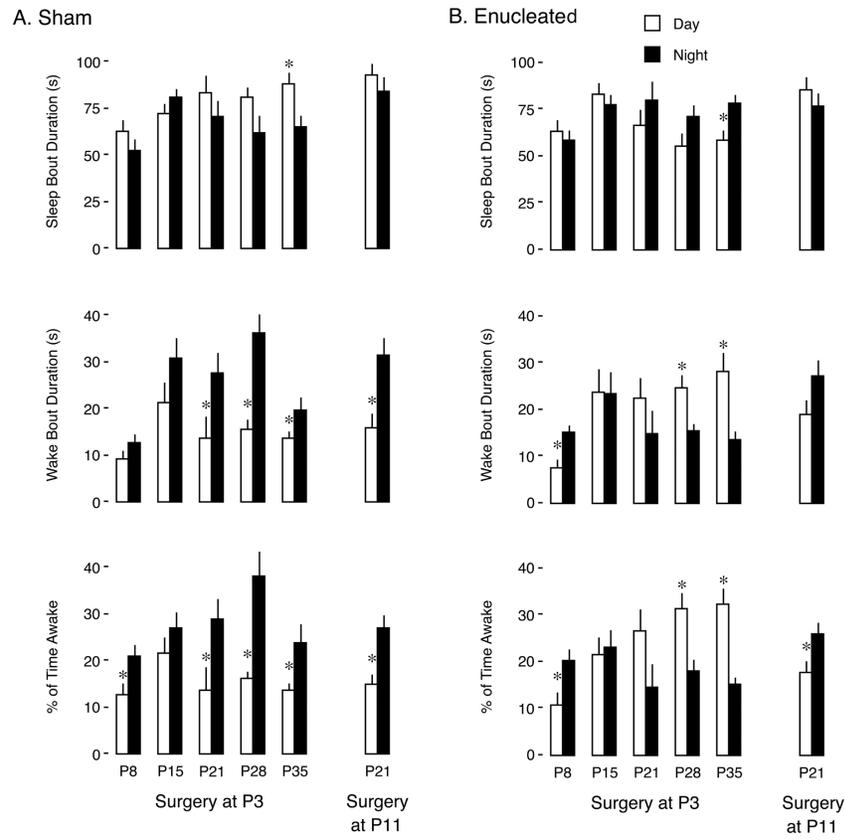


Figure 3. Day-night differences in sleep and wake parameters in sham and enucleated infant rats. Mean sleep bout durations (top row), wake bout durations (middle row), and percentage of time awake (bottom row) during the day and night in P8, P15, P21, P28, and P35 rats after sham surgery (A) or bilateral enucleation (B) at P3. Also shown are data from P21 rats after sham or enucleation surgery at P11. *Significantly different from corresponding nighttime value. $n = 6-8$ subjects per group. Means + SE.

after birth. Specifically, at P2, sleep and wake bout durations were generally shorter at night, and combined to result in approximately 50% more sleep-wake cycles at night than during the day. Nocturnal wakefulness first emerged around P15—at around the same time as did power-law wake behavior (Blumberg et al., 2005)—and became even more pronounced by P21. It was also at these ages that the number of sleep-wake cycles at night and during the day achieved balance (see Figure 1D), perhaps reflecting reciprocal circadian modulation of the SCN on downstream neural circuitry governing sleep and wakefulness.

Developmental studies in rats have shown that nearly all neurons in the SCN are formed by embryonic day (E)18 (Ifft, 1972) and, by E19, the SCN is more metabolically active during the day than at night and is synchronized to the dam's SCN activity (Reppert and Schwartz, 1984). Therefore, it is plausible that SCN

circadian activity, entrained to that of the dam, modulates sleep-wake rhythmicity as early as the 1st post-natal week.

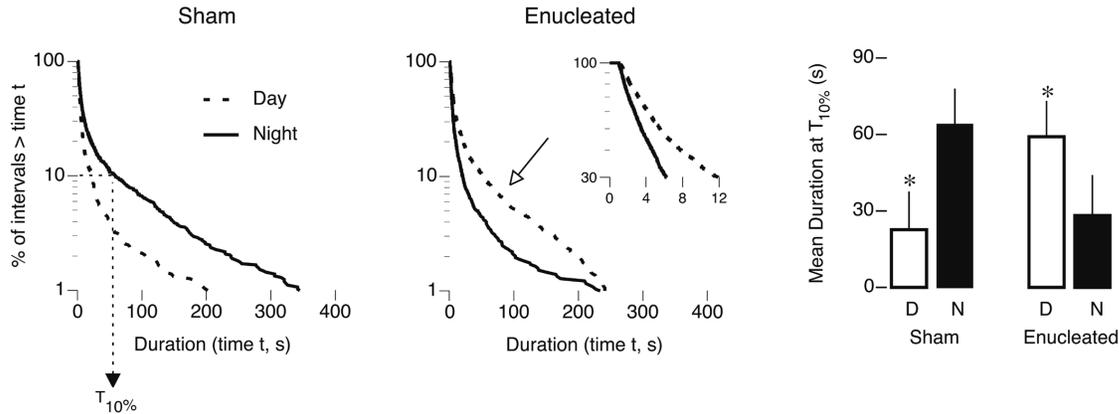
Constraints associated with the provisioning of milk may also play a role in shaping the development of circadian sleep-wake rhythms. The rat dam's milk ejection reflex only occurs during slow-wave sleep (Lincoln, 1983; Voloschin and Tramezzani, 1979). Consequently, pups suckle more, receive more milk, and gain more weight during the day than at night (Levin and Stern, 1975; Thiels et al., 1990). The transition to a nocturnal feeding pattern appears to occur soon after weaning and the transition to solid food (Levin and Stern, 1975).

In experiment 2, enucleation at P3 or P11 did not prevent the development of day-night differences in sleep-wake activity or power-law wake behavior. However, enucleation at P3 did result in P21 rats that exhibited longer wake bout durations during the day. Critically, because this pattern was sustained at P28 and P35, it is unlikely that these enucleated subjects were free-running. In contrast, enucleation at P11

resulted in P21 rats that exhibited longer wake bout durations at night, similar to shams (although it should be noted that testing these subjects only at P21 makes it more difficult to be confident that they were not free-running). These results suggest that enucleated infant rats differentially entrain to social zeitgebers within the nest environment and colony room depending on when the enucleation takes place (Mistlberger and Skene, 2004). Indeed, in pups blinded at birth and tested beyond the 1st postnatal week, nursing mothers set the phase of circadian SCN activity (Ohta et al., 2002), pineal serotonin N-acetyltransferase activity and corticosterone levels (Takahashi and Deguchi, 1983), and drinking (Shimoda et al., 1986) and locomotor (Honma et al., 1987) activity.

Because the SCN is more metabolically active during the day than at night in both diurnal and nocturnal species (Smale et al., 2003), we do not expect that enucleation in early infancy alters the circadian timing of

A. Surgery at P3



B. Surgery at P11

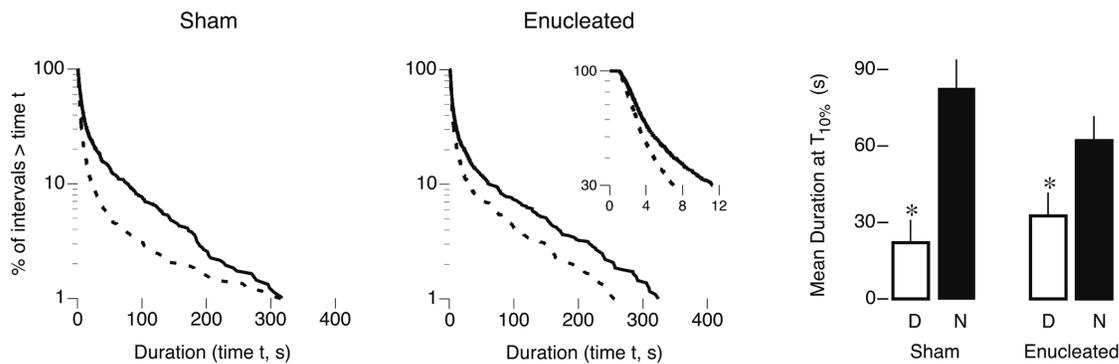


Figure 4. Effects of enucleation at P3 or P11 on wake bout durations during the day and night in P21 rats. (A) Log-survivor plots of pooled wake bout durations for P21 subjects that experienced sham surgery or enucleation on P3 (1339-1976 points per plot). Pups were tested during the day (dotted line) or at night (solid line). Open arrow indicates group exhibiting longer wake bout durations during the day. $T_{10\%}$ is indicated in the far-left plot to illustrate how this value was derived. Insets provide magnified views to reveal consistency of day-night differences in enucleated subjects even at short bout durations. The right panel presents mean values of $T_{10\%}$ across individual subjects. (B) Same as in (A) except pups experienced enucleation or sham surgery on P11 (897-1409 points per plot). *Significantly different from the corresponding nighttime value. Means + SE.

SCN metabolic activity. Rather, we hypothesize that visual system stimulation—from light and/or spontaneous activity within the retina—transmitted through the RHT to the SCN, induces functional changes in SCN interactions with its downstream neural structures.

In support of this hypothesis, we note that in non-enucleated rats, the shift from maternal to photic entrainment depends on the development of functional connectivity between the RHT and SCN, which occurs between P0 and P12 (Duncan et al., 1986; Hannibal and Fahrenkrug, 2004; Speh and Moore, 1993). Moreover, in adults, light may act as a zeitgeber by up-regulating growth factors (e.g., NGF1-A, BDNF) in the SCN (Allen and Earnest, 2005; Liang et al., 2000;

Tanaka et al., 1999). Such up-regulation of gene activity—seen during everyday entrainment in adults—could also, we suggest, play an inductive, organizational role during a sensitive period when the RHT is establishing its connections with the SCN. Interestingly, it was recently suggested that photoperiodic conditions during the perinatal period in hamsters can organize the adult immune system (Weil et al., 2006).

Future research should address more thoroughly the effects of early blindness on the expression of circadian sleep-wake rhythms and locomotor activity in adults. In addition, it will be important to examine the mechanisms mediating the developmental effects of enucleation on SCN functionality. For example, if,

as hypothesized here, photic stimulation via the RHT induces organizational changes in the connectivity of the efferent connections from the SCN, then those changes should be detectable as altered anatomical and physiological connections between the SCN and its target nuclei (Schwartz et al., 2004). If this hypothesis is supported by subsequent research, it would suggest one mechanism by which species differences in nocturnality and diurnality have evolved (Smale et al., 2003; Mrosovsky and Hattar, 2005).

ACKNOWLEDGMENTS

This work was supported by a research grant (MH50701) and an Independent Scientist Award (MH66424) from the National Institute of Mental Health (to M.S.B.). The second and third authors contributed equally to this project.

REFERENCES

- Allen GC and Earnest DJ (2005) Overlap in the distribution of TrkB immunoreactivity and retinohypothalamic tract innervation of the rat suprachiasmatic nucleus. *Neurosci Lett* 376:200-204.
- Blumberg MS, Seelke AM, Lowen SB, and Karlsson KA (2005) Dynamics of sleep-wake cyclicity in developing rats. *Proc Natl Acad Sci U S A* 102:14860-14864.
- Blumberg MS and Stolba MA (1996) Thermogenesis, myoclonic twitching, and ultrasonic vocalization in neonatal rats during moderate and extreme cold exposure. *Behav Neurosci* 110:305-314.
- Duncan MJ, Banister MJ, and Reppert SM (1986) Developmental appearance of light-dark entrainment in the rat. *Brain Res* 369:326-330.
- Ellison N, Weller JL, and Klein DC (1972) Development of a circadian rhythm in the activity of pineal serotonin N-acetyltransferase. *J Neurochem* 19:1335-1341.
- Frank MG and Heller HC (1997) Development of diurnal organization of EEG slow-wave activity and slow-wave sleep in the rat. *Am J Physiol* 273:R472-R478.
- Fuller PM, Gooley JJ, and Saper CB (2006) Neurobiology of the sleep-wake cycle: sleep architecture, circadian regulation, and regulatory feedback. *J Biol Rhythms* 21:482-493.
- Gramsbergen A (1976) The development of the EEG in the rat. *Dev Psychobiol* 9:501-515.
- Hannibal J and Fahrenkrug J (2004) Melanopsin containing retinal ganglion cells are light responsive from birth. *Neuroreport* 15:2317-2320.
- Henning SJ and Gisel EG (1980) Nocturnal feeding behavior in the neonatal rat. *Physiol Behav* 25:603-605.
- Honma K, Honma S, Shirakawa T, and Hiroshige T (1987) Phase setting of circadian locomotor rhythm of infant rats. *Am J Physiol* 252:R256-R261.
- Ifft JD (1972) An autoradiographic study of the time of final division of neurons in rat hypothalamic nuclei. *J Comp Neurol* 144:193-204.
- Karlsson KÆ and Blumberg MS (2005) Active medullary control of atonia in week-old rats. *Neuroscience* 130:275-283.
- Karlsson KÆ, Kreider JC, and Blumberg MS (2004) Hypothalamic contribution to sleep-wake cycle development. *Neuroscience* 123:575-582.
- Kittrell EMW and Satinoff E (1986) Development of the circadian rhythm of body temperature in rats. *Physiol Behav* 38:99-104.
- Levin R and Stern JM (1975) Maternal influences on ontogeny of suckling and feeding rhythms in the rat. *J Comp Physiol Psychol* 89:711-721.
- Liang FQ, Allen G, and Earnest D (2000) Role of brain-derived neurotrophic factor in the circadian regulation of the suprachiasmatic pacemaker by light. *J Neurosci* 20:2978-2987.
- Lincoln DW (1983) Physiological mechanisms governing the transfer of milk from mother to young. In *Symbiosis in Parent-Offspring Interactions*, Rosenblum LA and Moltz H, eds, pp 77-112, New York, Plenum.
- Mistlberger RE and Skene DJ (2004) Social influences on mammalian circadian rhythms: animal and human studies. *Biol Rev Camb Philos Soc* 79:533-556.
- Moore RY (1983) Organization and function of a central nervous system circadian oscillator: the suprachiasmatic hypothalamic nucleus. *Fed Proc* 42:2783-2789.
- Mrosovsky N and Hattar S (2005) Diurnal mice (*Mus musculus*) and other examples of temporal niche switching. *J Comp Physiol [A]* 191:1011-1024.
- Nuesslein-Hildesheim B, Imai-Matsumura K, Döring H, and Schmidt I (1995) Pronounced juvenile circadian core temperature rhythms exist in several strains of rats but not in rabbits. *J Comp Physiol B* 165:13-17.
- Ohta H, Honma S, Abe H, and Honma K (2002) Effects of nursing mothers on rPer1 and rPer2 circadian expressions in the neonatal rat suprachiasmatic nuclei vary with developmental stage. *Eur J Neurosci* 15:1953-1960.
- Reppert SM and Schwartz WJ (1984) The suprachiasmatic nuclei of the fetal rat: characterization of a functional circadian clock using ¹⁴C-labeled deoxyglucose. *J Neurosci* 4:1677-1682.
- Reppert SM, Weaver DR, and Rivkees SA (1988) Maternal communication of circadian phase to the developing mammal. *Psychoneuroendocrinology* 13:63-78.
- Rusak B and Zucker I (1979) Neural regulation of circadian rhythms. *Physiol Rev* 59:449-526.
- Schwartz MD, Nunez AA, and Smale L (2004) Differences in the suprachiasmatic nucleus and lower subparaventricular zone of diurnal and nocturnal rodents. *Neuroscience* 127:13-23.
- Seelke AM, Karlsson KA, Gall AJ, and Blumberg MS (2005) Extraocular muscle activity, rapid eye movements and the development of active and quiet sleep. *Eur J Neurosci* 22:911-920.
- Sekaran S, Lupi D, Jones SL, Sheely CJ, Hattar S, Yau KW, Lucas RJ, Foster RG, and Hankins MW (2005) Melanopsin-dependent photoreception provides earliest light detection in the mammalian retina. *Curr Biol* 15:1099-1107.

- Shimoda K, Hanada K, Yamada N, Takahashi K, and Takahashi S (1986) Periodic exposure to mother is potent zeitgeber of rat pups' rhythm. *Physiol Behav* 36:723-730.
- Smale L, Lee T, and Nunez AA (2003) Mammalian diurnality: some facts and gaps. *J Biol Rhythms* 18:356-366.
- Speh JC and Moore RY (1993) Retinohypothalamic tract development in the hamster and rat. *Brain Res Dev Brain Res* 76:171-181.
- Spiers DE (1988) Nocturnal shifts in thermal and metabolic responses of the immature rat. *J Appl Physiol* 64:2119-2124.
- Takahashi K and Deguchi T (1983) Entrainment of the circadian rhythms of blinded infant rats by nursing mothers. *Physiol Behav* 31:373-378.
- Tanaka M, Iijima N, Amaya F, Tamada Y, and Ibata Y (1999) NGFI-A gene expression induced in the rat suprachiasmatic nucleus by photic stimulation: spread into hypothalamic periventricular somatostatin neurons and GABA receptor involvement. *Eur J Neurosci* 11:3178-3184.
- Thiels E, Alberts JR, and Cramer CP (1990) Weaning in rats: II. Pup behavior patterns. *Dev Psychobiol* 23:495-510.
- Voloschin LM and Tramezzani JH (1979) Milk ejection reflex linked to slow wave sleep in nursing rats. *Endocrinology* 105:1202-1207.
- Weil ZM, Pyter LM, Martin LB 2nd, and Nelson RJ (2006) Perinatal photoperiod organizes adult immune responses in Siberian hamsters (*Phodopus sungorus*). *Am J Physiol Regul Integr Comp Physiol* 290:R1714-R1719.